



***In vitro* CONTROL OF FISH HELMINTH PARASITE,  
*Euclinostomum heterostomum* BY FLAVONOID-RICH  
EXTRACTS OF *Mangifera indica***

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**AUTHORS' CONTRIBUTIONS**

This work was carried out in collaboration among all authors. Author IK gave the concept, designed the study, did the analysis or interpretation and wrote the manuscript. Authors SR, YK and MMR did the data collection or processing and managed the literature search. All authors read and approved the final manuscript.

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**ABSTRACT**

The study was conducted to determine anthelmintic activity of mango plant (*Mangifera indica*) known widely for its medicinal value. Alcoholic extracts of leaves of mango plant was tested as a therapeutic agent for controlling *Euclinostomum heterostomum*, trematode parasite in fish by 'in vitro' method. Qualitative and quantitative estimation of the ethyl acetate extract showed higher flavonoid content compared to ethanol and chloroform extracts. The ethyl acetate extract was found to be effective in killing parasites significantly (LC<sub>50</sub> value 8.2016 mg/l) which clearly demonstrates the anthelmintic activity of mango leaves. Acute toxicity tests of ethyl acetate extracts on host animals (*Channa punctata*) showed the lethal concentration (LC<sub>50</sub>) value of 194.98 mg/l, thus a considerable lower effective working solution can be used for application for control of parasites without causing damage to host animals. On the basis of above study, it can be concluded that the mango leaves can be used as an alternative to synthetic drugs applied indiscriminately to control helminth parasites which has resulted in serious health concern.

**Keywords:** Anthelmintic; *Channa punctata*; trematode; mango.

**1. INTRODUCTION**

In India intensification of aquaculture practices has resulted in wide spread outbreak of parasites and microorganisms causing a serious public health issues [1]. Indiscriminate use of synthetic drugs and chemicals has lead to multidrug resistance, bioaccumulation, bioconcentration and transmission of drug residues in the food chain. Hence search for alternative modes for control of parasites has lead to the use of plant based herbal drugs.

Plants are known to possess secondary metabolites like flavonoids, alkaloids, terpenoids, steroids, phenols, glycosides and tannins which are made of several active components accounting to the therapeutic effects of the plants [2]. Flavonoids are also known as nature's tender drugs as they possess various pharmacological activities making them a huge reservoir of medicines and drugs [3,4].

Medicinal plants like mango are known to treat various diseases. Mango tree (*Mangifera indica* L.),

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belonging to Anacardiaceae are widely distributed in India and are known to possess antimicrobial [5], antifungal [6], antitumor [7] activities.

*Mangifera indica* has been known for its ethnopharmacological value and has been used for a long time as an anthelmintic agent [8,9]. Patel et al. [10] has worked on anthelmintic activity of *Mangifera indica* leaves using *Pheretima posthuma* as model in general however reports on efficacy of its secondary metabolites, antioxidant activity was not reported.

Hence the present study was carried to evaluate the efficiency of flavonoid-rich fractions of mango leaves in controlling helminthiasis in fish using *Euclinostomum heterostomum* [11] as a model (helminth) for *in vitro* evaluation to justify the traditional beliefs.

## 2. MATERIALS AND METHODS

### 2.1 Study Location for Host Sampling and Collection of Helminthes

The study was conducted on 240 specimens of *Channa punctata* (length 12-17 cm and weight 40-60 g) reared in fish farms under the tropical conditions of West Bengal. The trematodes collected from *Channa punctata* were kept in 0.67% saline solution in a petridish incubated at 37°C for use in *in vitro* studies. Few were fixed with AFA or 70% ethanol. Morphometric identification was performed using selected identification keys [12,13,14].

### 2.2 Qualitative Evaluation of Plant Extracts

Dried leaves of mango plant was prepared for qualitative evaluation [15]. Detection of flavonoids (Alkaline Reagent Test), alkaloids (Mayer's Test: and Dragendroff's Test) [16], and saponins (Froth Test) [17] was performed. Test for steroids and triterpenoids (Salkowski test), phenols (Ferric Chloride Test) [18] and tannins (Gelatin Test) was performed [19].

### 2.3 Quantitative Estimation of Secondary Metabolites

Dried leaves of mango plant was prepared according to [15] for quantitative evaluation. Total phenolic content of ethanol, chloroform, ethyl acetate extract of mango leaves was determined with Folin-Ciocalteu method [20] using Gallic acid was used as the reference compound. Total flavonoid content of the mango leaves were determined according to a modified colorimetric method [21] using Quercetin as standard. 1,1-diphenyl-2-picrylhydrazine (DPPH)

radical scavenging activity was performed according to [22]. Half maximal inhibitory concentration (IC<sub>50</sub>) values denote the concentration of sample which is required to scavenge 50% of DPPH free radicals.

### 2.4 Screening for *In-vitro* Anthelmintic Assay

*In vitro* bioassay was carried out using groups of trematodes (4-5 each) in which 25 ml of various concentration of ethyl acetate extracts and Albendazole as standard was prepared in a Petri dish [23]. Observations were made from the time taken for paralysis of trematodes to set-in was noted. Control worms were maintained in phosphate-buffered saline with 1% dimethylsulfoxide at (37±1)°C.

### 2.5 Acute Toxicity Test and Estimation of LC<sub>50</sub> of Experimental Fishes

Five day bioassay was performed to determine the acute toxicity of ethyl acetate extracts using varying concentrations (10, 25, 50, 75 and 100 mg/l) on host animal [24]. The LC<sub>50</sub> results were analyzed according to Probit analysis [25] and the working dose was determined. All treatments were done in three replicates.

### 2.6 Statistical Analysis

Data was represented as mean ± SD for each group (n=6). Statistical significance, SD and analysis of variance (ANOVA) was determined at 5% level. P<0.005 was considered significant.

## 3. RESULTS AND DISCUSSION

Helminthes are the most widespread parasitic group infecting both invertebrate and vertebrate. In the present study a total of 68 parasites representing different species and some unidentified species were collected from the host fish (*Channa punctata*). The parasite predominantly obtained from the liver of the infected fishes was identified as *Euclinostomum heterostomum*, a common trematode parasite [11].

Indiscriminate use of commercially available anthelmintic drugs like ampicillin, cefotaxime, streptomycin, kanamycin, gentamicin have resulted in resistance of parasites [26] thus there is an urgent need for alternative modes for controlling this problem. The phytochemicals present in plants can act as active constituent for controlling the rate of parasitic infestation. The preliminary phytochemical analysis of ethyl acetate extracts of mango leaf powder exhibited higher number of secondary metabolites like, triterpenoids, alkaloids, flavanoids, phenols, saponins and tannins in comparison to ethanolic and chloroform extract (Table 1).

**Table 1. Phytochemical constituents of *Mangifera indica* extracts based on solvents used**

S/No.	Phytochemical constituents	Various extract of plant material		
		Chloroform	Ethyl acetate	Ethanol
1.	Flavonoids	+	+++	++
2.	Alkaloids	-	++	+
3.	Steroids	+	-	-
4.	Tannins	+	+	+
5.	Saponins	-	+	-
6.	Triterpenoids	+	++	-
7.	Glycosides	-	-	+++
8.	Phenols	+	+	+

(Present: +; Absent: -; ++moderate presence; +++high presence)

The concentrations of total phenolic contents and of flavonoids (Quantitative assay) in the ethyl acetate extracts was found to be higher than in chloroform and ethanolic extracts (Tables 2 and 3). Flavonoids are known for its antiparasitic effects against a number of parasites and microbes [27,28,29]. The DPPH scavenging activity is shown in Fig 1. The radical scavenging effects of the extracts showed concentration-dependence. Ascorbic acid was used as a standard antioxidant as it has strong DPPH scavenging property [30].

Udem et al. [31] reported that the *Mangifera indica* ethanolic extract showed  $79.09 \pm 0.42$  percentage inhibition at 100 mg/ml while 20 mg/ml showed  $73.17 \pm 1.81$  percentage inhibition in DPPH scavenging activity while in the present study chloroform extract, ethyl acetate and ethanol at 100  $\mu\text{g/ml}$  showed  $73.37 \pm 1.25$ ,  $94.3 \pm 2.34$  and  $81.7 \pm 3.25$  percentage inhibition which is much higher than the earlier study.

The  $\text{IC}_{50}$  values in the present study were found to be  $59.075 \pm 2.21 \mu\text{g/ml}$ ,  $49.169 \pm 0.11 \mu\text{g/ml}$ ,  $24.366 \pm 1.88 \mu\text{g/ml}$ ,  $6.2341 \pm 3.34 \mu\text{g/ml}$  for chloroform, ethanol, ethyl acetate and ascorbic acid respectively. The lower the  $\text{IC}_{50}$  values the higher the free radical activity. Hence it can be concluded that ethyl acetate fractions have higher antioxidant activity than chloroform and ethanol extracts. Thus the role of flavonoids and antioxidant properties of *Mangifera indica* for control of helminth parasites in fishes have been studied by *in vitro* method.

**Table 2. Concentrations of total phenolic content in the plant extracts expressed in terms of Gallic acid equivalent (GAE) (mg of QE/g of extract)**

Extract	Milligram of GAE/g of plant extract
Chloroform Extract	108.12
Ethyl Acetate extract	360.10
Ethanolic extract	258.16

**Table 3. Concentrations of flavonoids in the plant extracts expressed in terms of Quercetin equivalent (QE) (mg of QE/g of extract)**

Extract	Milligram of GAE/gm of plant extract
Chloroform Extract	108.12
Ethyl Acetate extract	360.10
Ethanolic extract	258.16

*In vitro* tests to evaluate the worm motility for prospecting of novel anthelmintic agents was reported [32,33]. The advantage of these assays are that the compounds or extracts are tested with the different life-cycle stages of the parasite in direct contact. So far, *in vitro* screenings of potential anthelmintic agents prior to *in vivo* testing have been shown to be a rational and practical strategy since they save time and money and also minimize the number of animals necessary for the development of a new therapeutic agent [34].

Thus *in vitro* assay for evaluating effect of ethyl acetate extract of *Mangifera indica* against *Euclinostomum heterostomum* was performed on varying doses for twenty four hours [23]. The results (Table 4), showed a significant dose-response relationship, with higher doses causing greater mortality. After twenty four hr exposure ethyl acetate extract at concentrations of 10 mg/l or more resulted in 60% mortality. Anthelmintic activity of plant extracts on *E. heterostomum* reported in present study is in agreement with the findings of earlier workers on different helminth parasites [35-41].

The lethal concentration ( $\text{LC}_{50}$ ) value of the different doses was calculated arithmetically using probit analysis [25]. The lethal concentration ( $\text{LC}_{50}$ ) values of ethyl acetate extract of *Mangifera indica* was determined and found to be 8.2016 mg/l (Table 5 and Fig. 2). Tariq et al. [37] reported  $\text{LC}_{50}$  for aqueous and ethanolic extracts of *Achillea millifolium* on *Haemonchus contortus* to be 0.05 and 0.11 mg/ml

respectively whereas Aggarwal et al. [42] showed that  $LC_{50}$  values to be  $12.05 \text{ mg/ml} \pm 3.24$  and  $23.52 \text{ mg/ml} \pm 6.4$  for *C. procera*,  $24.37 \text{ mg/ml} \pm 4.11$  and  $21.02 \text{ mg/ml} \pm 4.6$  for *A. indica*,  $18.92 \text{ mg/ml} \pm 4.54$  and  $24.43 \text{ mg/ml} \pm 6.96$  for *P. granatum* for ethanolic

and aqueous extracts respectively. Different ranges of concentration and efficacy were observed by various researchers working with different solvents, extraction procedures for isolation of active molecules which corroborates with the present study [43,44].

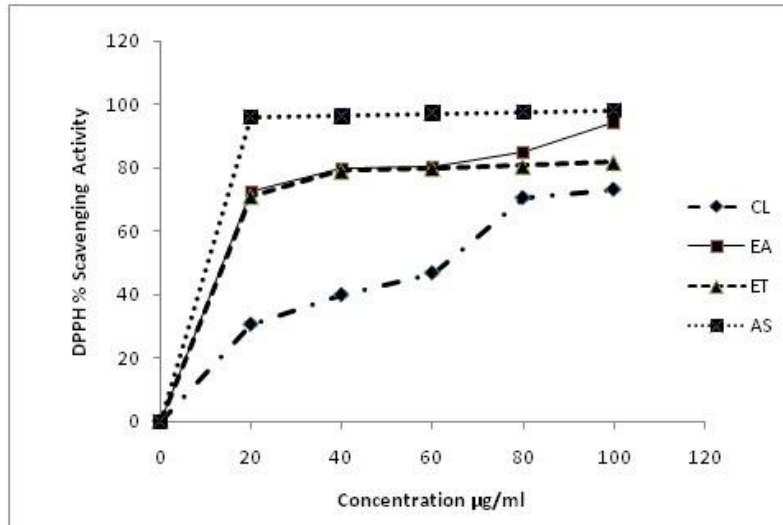


Fig. 1. DPPH scavenging activity assay of various extracts of *Mangifera indica*  
CL-Chloroform, EA-Ethyl Acetate, ET-Ethanol, AS-Ascorbic acid

Table 4. Survivability of parasites in ethyl acetate extract of *Mangifera indica*

Dose (mg/l)	Time of paralysis (hour)	Time of death(hour)
1	15 ± 2.1	28 ± 1.5
2	12 ± 0.32	22 ± 0.85
4	10 ± 2.36	18 ± 3.21
6	7 ± 1.98	16 ± 2.47
8	5 ± 0.25	14 ± 2.15
10	3 ± 2.69	12 ± 1.28
12	2 ± 0.37	10 ± 1.32
14	1.5 ± 1.57	9 ± 5.48
16	0.75 ± 8.23	7 ± 0.27
Standard	3.5 ± 0.65	13 ± 1.2

Table 5. Acute toxicity of ethyl acetate extracts of *Mangifera indica* to trematode parasite

Concentration (mg/l)	Log10 concentration	Total number of animals	Number of animals dead	% Mortality	Corrected mortality (%)	Probit
0	0	5	0	0	0	-
1	0	5	0	0	0	-
2	0.3001	5	1	20	20	4.16
4	0.60206	5	1	20	20	4.16
6	0.778151	5	2	40	40	4.75
8	0.90309	5	2	40	40	4.16
10	1	5	3	60	60	5.25
12	1.0792	5	3	60	60	5.25
14	1.1461	5	4	80	80	5.84
16	1.2041	5	4	80	80	5.84

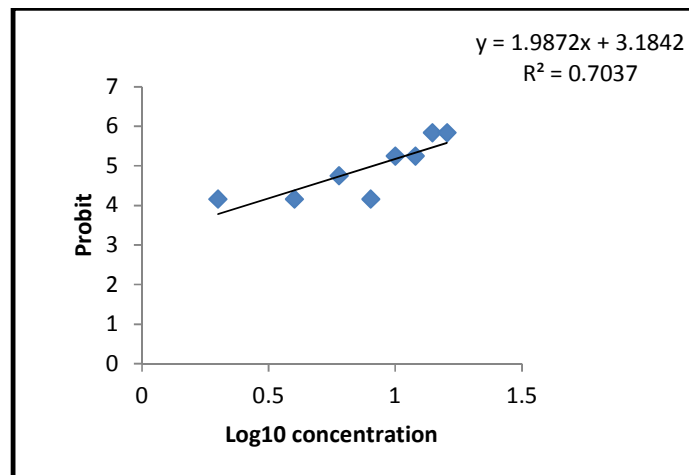


Fig. 2. Probit analysis for estimation of  $LC_{50}$

The use of crude extracts as anthelmintic drugs is essential to study if the extract has any kind of toxic effects over healthy fish due to its application. A bioassay was hence performed for five days for determining the acute toxicity of ethyl acetate extracts of *Mangifera indica* to healthy fishes and lethal concentration ( $LC_{50}$ ) was estimated (Table 6 and Fig. 3). The fish survived easily in the ethyl acetate extract at doses of 10–75 mg/l for five days (Fig. 3). However on increasing concentration reduced the survivability of the fishes which might be due to suspension of solids over gills thus reducing the oxygen supply to the gills. Hence increasing the concentration of the extracts up to 75 mg/l does not have any detrimental effect

on the host and can be effectively used in aquaculture.

The  $LC_{50}$  value was found to be 194.98 mg/l which was much higher than the effective working solutions ie 10–75 mg/l used in the experiments (Table 6 and Fig. 4). Thus ethyl acetate extracts can effectively be used in controlling helminth parasites in aquaculture without causing any detrimental effect on fishes.  $LC_{50}$  values of the distilled water, 50% ethyl alcohol and absolute ethyl alcohol extracts of *A. zapota* leaf parts for *C. punctatus* were 279.35, 175.89, 125.69 ppms, respectively were reported by Nasiruddin et al. [45]. Similar results were observed by various workers in terms of the working with plant extracts [46–48].

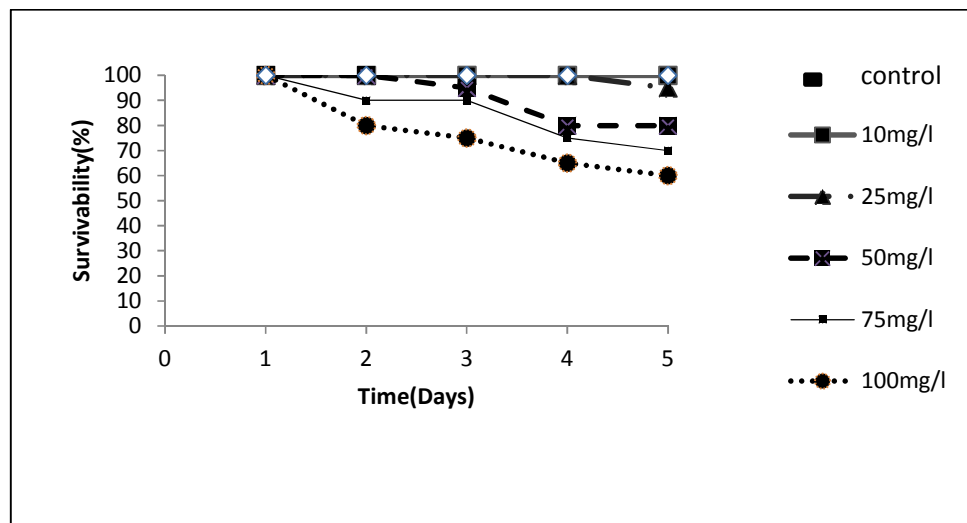
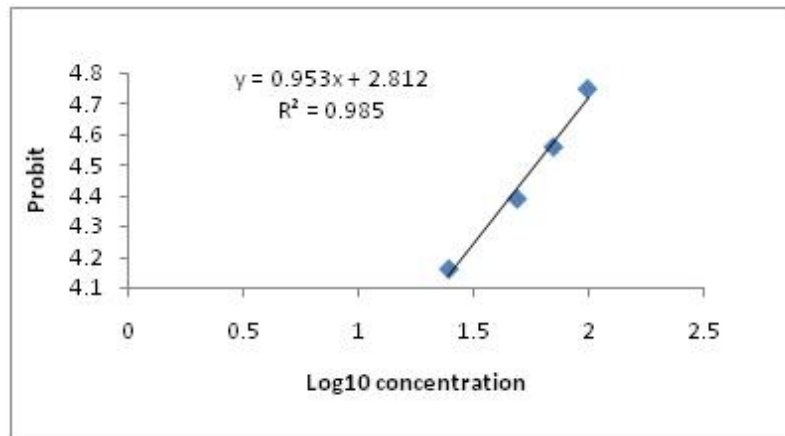


Fig. 3. Graphical representation of the survivability % of fish during acute toxicity of ethyl acetate extracts of mango to fish after 5 days of bioassay

**Table 6. Acute toxicity of ethyl acetate extracts of mango to fish after 5 days of bioassay**

Concentration (mg/l)	Log <sub>10</sub> concentration	Total number of animals	Number of animals dead	% Mortality	Corrected mortality (%)	Probit
Control	0	15	1	6.67	0	-
10	1	15	0	0	0	-
25	1.39	15	4	26.67	20	4.16
50	1.69	15	5	33.33	26.68	4.39
75	1.85	15	6	40	33.35	4.56
100	2	15	7	46.67	40.02	4.75

**Fig. 4. Probit analysis for estimation of LC<sub>50</sub> values**

#### 4. CONCLUSION

The present study using mango leaves extract inhibited motility of trematode parasites at low concentration, increasing the concentration of the plant extracts resulted in early paralysis, thus indicating dose-dependent efficacy of the test materials. Similar observation of using plant extract as anthelmintic drug has been made by some authors [49-52]. Moreover low toxic effect on host fishes as evaluated from LC<sub>50</sub> values make it an effective for its use in aquaculture practices.

These findings indicated that the mango leaves extract contains active principles which can act as effective drugs against the trematode parasites affecting the fish. Hence it is of utmost necessity to identify bioactive compounds present in mango leaves which might be responsible for its anthelmintic activity.

#### ETHICS APPROVAL

As per CPCSEA instruction's protocol for experimentation on fishes, does not require approval from ethical committee.

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#### COMPETING INTERESTS

Authors have declared that no competing interests exist.

#### REFERENCES

1. Walker PJ, Winton JR. Emerging viral diseases of fish and shrimp. *Vet Res.* 2010;41:51.
2. Arunkumar S, Muthuselvam M. Analysis of phytochemical constituents and antimicrobial activities of *Aloe vera* L. against clinical pathogens. *World J Agric Res.* 2009;5:572-576.
3. Kumar S, Pandey AK. Chemistry and biological activities of flavonoids: An overview. *Sci World J.* 2013;162750. DOI: 10.1155/2013/162750
4. Działo M, Mierziak J, Korzun U, Preisner M, Szopa J, Kulma A. The potential of plant

- phenolics in prevention and therapy of skin disorders. *Int J Mol Sci*. 2016;17:160.  
DOI: 10.3390/ijms17020160
5. Durapandiyar V, Ayyanar M, Ignacimuthi S. Antimicrobial activity of some ethnomedicinal plants used by Paliyar from Tamil Nadu, India. *BMC Complem Altern Med*. 2006;6: 35.  
DOI: 10.1186/1472-6882-6-35
6. Stoilova I, Gargova S, Stoyanova L Ho. Antimicrobial and antioxidant activity of the polyphenol mangiferin. *Herba Polonica*. 2005; 51:37-44.
7. Paria ND. Medicinal plant resources of South West-Bengal. Kolkata: Directorate of forests, Government of West Bengal. 2005;39-40.
8. Telang M, Dhulap S, Mandhare A, Hirwani R. Therapeutic and cosmetic applications of mangiferin: A patent review, Expert Opinion on Therapeutic Patents. 2013;23(12):1561-1580.
9. Jyotshna Khare P, Shanker K. Mangiferin. A review of sources and interventions for biological activities *Bio Factors*. 2016;42(5): 504–514
10. Patil D, Halle P, Bade A. *In-vitro* anthelmintic activity of methanolic extract of *Mangifera indica* leaves. *World J Pharm Pharmac Sci*. 2014;12(3):771-776.
11. Caffara M, Locke SA, Cristanini C, Davidovich N, Markovich MP, Fioravanti ML. A combined morphometric and molecular approach to identifying metacercariae of *Euclinostomum heterostomum* (Digenea: Clinostomidae). *J Parasitol*. 2016;102(2):239-48.  
DOI: 10.1645/15-823
12. Yamaguti S. *Systema helminthum*. The digenetic trematodes of vertebrates Part I and II. New York. 1958;1:1141–1143.
13. Moravec F. Parasitic nematodes of freshwater fishes of Europe. Kluwer Academic Publishers, Dordrecht/Boston/London; 1994.
14. Anderson RC. Nematode parasites of vertebrates their development and transmission. 2<sup>nd</sup> Edition, CABI Publishing; 2000.
15. Jiménez E, Dorta F, Medina C, Ramírez A, Ramírez I, Peña-Cortés H. Anti-phytopathogenic activities of macro-algae extracts. *Mar Drugs*. 2011;9:739-756.  
DOI: 10.3390/md9050739
16. Siddiqui AA, Ali M. Practical pharmaceutical chemistry. First edition. CBS Publishers and distributors, New Delhi. 1997;126–131.
17. Ejikeme CM, Ezeonu CS, Eboatu AN. Determination of physical and phytochemical constituents of some tropical timbers indigenous to Niger Delta Area of Nigeria. *Eur Sci J*. 2014;10(18):247–27.
18. Debela A. Manual for phytochemical screening of medicinal plants. Ethiopian Health and Nutrition Research Institute, Addis Ababa. Ethiopia. 2002;35–47.
19. Iyengar MA. Study of drugs. Eighth edition. Manipal Power Press, Manipal, India; 1995.
20. Singleton VL, Orthofer R, Lamuela-Raventos RM. Analysis of total phenols and other oxidation substrates and antioxidants by means of Folin-Ciocalteu reagent. *Methods Enzymol*. 1999;299:152-178.
21. Lee WH, Intan SI. Antioxidant activity, total phenolics and total flavonoids of *Syzygium polyanthum* (Wight) walp leaves. *Int J Med Arom Plnt*. 2012;2(2):219-228.
22. Blois MS. Antioxidant determination by the use of a stable free radical nature. *Nature*. 1958;26:1199-1200.
23. Agarwal K, Singh H. An *in-vitro* evaluation of anthelmintic activity of *Morus alba* bark. *MIT Int J Pharm Sci Res*. 2016;2(2):35–37
24. Chromcova L, et al. NeemAzal T/S – toxicity to early-life stages of common carp (*Cyprinus carpio* L.). *Vet Med – Czech*. 2015;60(1):23–30
25. Finney DJ. Probit analysis. 3<sup>rd</sup> Edition, Cambridge University Press, Cambridge. 1971; 2526.
26. Romero J, Carmen F, Navarrete P. Antibiotics in aquaculture – Use, abuse and alternatives; 2012.  
DOI: 10.5772/28157
27. Sharma SK, Parasuraman P, Kumar G, Surolia N, Surolia A. Green tea catechins potentiate triclosan binding to enoyl-ACP reductase from *Plasmodium falciparum* (PfENR). *J Med Chem* 2007;50:765–775.  
DOI: 10.1021/jm061154d
28. Soto J, Gómez C, Calzada F, Ramírez ME. Ultrastructural changes on *Entamoeba histolytica* HM1-IMSS caused by the flavan-3-ol, (-)-epicatechin. *Planta Med*. 2010;76:611–612.

- DOI: 10.1055/s-0029-1240599
29. Dodson HC, Lyda TA, Chambers JW, Morris MT, Christensen KA, Morris JC. Quercetin, a fluorescent bioflavonoid, inhibits *Trypanosoma brucei* hexokinase. *Exp Parasitol*. 2011;127: 423–428.  
DOI: 10.1016/j.exppara.2010.10.011
30. Tona L, Kambu K, Ngimbi N, Cimanga K, Vlietinck AJ. Antiamoebic and phytochemical screening of some Congolese medicinal plants. *J Ethnopharmacol*. 1998; 61(1):57–65.
31. Udem CG, Dahiru D, Etteh CC. *In vitro* antioxidant activities of aqueous and ethanol extracts of *Mangifera indica* leaf, stem-bark and root-bark. *Commn*. 2018;8(3):119-124.
32. Costa CTC, Morais SM, Bevilaqua CML, Souza MMC, Leite FKA. Ovicidal effect of *Mangifera indica* L. seeds extracts on *Haemonchus contortus*. *Rev Bras Parasitol Vet*. 2002;11:57–60.
33. Vasconcelos ALC, Bevilaqua CM, Morais SM, Maciel MV, Costa CT, Macedo IT, Oliveira LM, Braga RR, Silva RA, Vieira LS. Anthelmintic activity of *Croton zehntneri* and *Lippia sidoides* essential oils. *Vet Parasitol*. 2007;148:288–294.
34. Ferreira LE, Castro PMN, Chagas ACS, França SC, Belebony RO. *In vitro* anthelmintic activity of aqueous leaf extract of *Annona muricata* L. (Annonaceae) against *Haemonchus contortus* from sheep. *Exp Parasitol*. 2013;134:327–32.
35. Ghosh NK, Babu SP, Sukul NC, Ito A. Cestocidal activity of *Acacia auriculiformis*. *J Helminthol*. 1996;70:171–172.
36. El Garhy MF, Mahmoud LH. Anthelmintic efficacy of traditional herbs on *Ascaris lumbricoides*. *J Egyptian Soc Parasitol*. 2002; 32:893–900.
37. Tariq KA, Chishti MZ, Ahmad F, Shawl AS. Anthelmintic efficacy of *Achillea millefolium* against gastrointestinal nematodes of sheep: *In vitro* and *in vivo* studies. *J Helminthol*. 2008; 82:227–233.
38. Lalchandama K, Roy B, Dutta B. Anthelmintic activity of *Acacia oxyphylla* stem bark against *Ascaridia galli*. *Pharm Biol* 2009;47:578-583.  
DOI: 10.1080/13880200902902463
39. Roy B, Swargiary A, Syiem D, Tandon V. *Potentilla fulgens* (family Rosaceae), a medicinal plant of north-east India: a natural anthelmintic. *J Parasit Dis*. 2010; 34(2):83–88.
40. Fouche G, Sakong BM, Adenubi OT, Pauw E, Leboh, T, Wellington KW, Eloff JN. Anthelmintic activity of acetone extracts from South African plants used on egg hatching of *Haemonchus contortus*. *Onderstepoort J Vet Res*. 2016;83(1):e1e7.  
DOI: doi.org/10.4102/ojvr.v83i1.1164
41. Lubian C, et al. Anthelmintic activity of plant aqueous extracts against *Panagrellus redivivus in vitro*. *Arq Inst Biol*. 2019;86:e0672018.  
Available:https://dx.doi.org/10.1590/1808-1657000672018
42. Aggarwal R, Kaur K, Suri M, Bagai U. Anthelmintic potential of *Calotropis procera*, *Azadirachta indica* and *Punica granatum* against *Gastrothylax indicus*. *J Parasit Dis*. 2016;40(4):1230-1238.
43. Costa CTC, Bevilaqua CML, Camurca-Vasconcelos ALF, Maciel MV, Morais SM, Castro CMS, Braga RR, Oliveira LMB. *In vitro* ovicidal and larvicidal activity of *Azadirachta indica* extracts on *Haemonchus contortus*. *Small Rumin Res*. 2008;74(1–3): 284–287.
44. Suteky T, Dwatmadji T. Anthelmintic activity of *Melastoma malabatricum* extract on *Haemonchus contortus* activity *in vitro*. *Asian J Pharm Clin Res*. 2011;4(1):68–70.
45. Nasiruddin M, Azadi M, Chakma D. Toxicological properties of *Achras zapota* (Linn) plant parts on the predatory fishes *Heteropneustes fossilis* (Bloch) and *Channa Punctatus* (Bloch). *Bangladesh J Zool*. 2012;40.  
DOI: 10.3329/bjz.v40i1.12900
46. Gholipour KH, Sahandi JTA. Influence of garlic (*Allium sativum*) and mother worth (*Matricaria chamomilla*) extract on *Ichthyophthirius multifiliis* parasite treatment in sail fin molly (*Poecilia latipinna*) ornamental fish. *APCBEE Procedia*. 2012;4:6–11.
47. Fu YW, Zhang QZ, Xu DH, Wang B, Liang JH, Lin DJ. Cynatratoside-C efficacy against theronts of *Ichthyophthirius multifiliis* and toxicity tests on grass carp and mammal blood cells. *Dis Aquat Org*. 2015;117(1):13–20.
48. Purivirojkul W. Potential application of extracts from Indian almond (*Terminalia catappa* Linn.) leaves in Siamese fighting fish (*Betta splendens* Regan) culture. *Commun Agric Appl Biol Sci*. 2012;77(4):439–448.
49. Dasgupta S, Roy B, Tandon V. Ultrastructural alterations for the tegument of *Raillietina echinobothrida* treated with stem bark of



- Acacia oxyphylla (Leguminosae). J Ethnopharmacol. 2010;127:568-571.
50. Abdel-Ghaffar F, Semmler M, Al-Rasheid SB, Strassen B, Fischer K, Aksu G, et al. The effects of different plant extracts on intestinal nematodes and trematodes. Parasitol Res. 2011;108:979-984.
  51. Ferriera JF, Peaden P, Keiser J. *In vitro* trematocidal effects of crude alcoholic extracts of *Artemisia annua*, *A. absinthium*, *Asiminatriloba*, and *Fumaria officinalis*: Trematocidal plant alcoholic extracts. Parasitol Res. 2011;109:1585-92.
  52. Hossain E, Chandra G, Nandy AP, Mandal SC, Gupta JK. Anthelmintic effect of a methanol extract of *Bombax malabaricum* leaves on *Paramphistomum explanatum*. Parasitol Res. 2012;110(3):1097-1102.